

Human alpha 1 Antichymotrypsin ELISA Kit is an Enzyme Immunoassay kit for the quantification of Human alpha 1 Antichymotrypsin in serum and plasma.

Catalog number: ARG82246

Package: 96 wells

For research use only. Not for use in diagnostic procedures.

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INTRODUCTION

Alpha 1-antichymotrypsin (symbol α 1AC, A1AC, or a1ACT) is an alpha globulin glycoprotein that is a member of the serpin superfamily. In humans, it is encoded by the SERPINA3 gene.

Alpha 1-antichymotrypsin inhibits the activity of certain enzymes called proteases, such as cathepsin G that is found in neutrophils, and chymases found in mast cells, by cleaving them into a different shape or conformation. This activity protects some tissues, such as the lower respiratory tract, from damage caused by proteolytic enzymes.

This protein is produced in the liver and it is an acute phase protein that is induced during inflammation. [Provide by Wikipedia: Alpha 1 Antichymotrypsin]

PRINCIPLE OF THE ASSAY

This assay employs the quantitative sandwich enzyme immunoassay technique. A capture antibody specific for Alpha 1 Antichymotrypsin has been pre-coated onto a microtiter plate. Standards or samples are pipetted into the wells and any Alpha 1 Antichymotrypsin present is bound by the immobilized antibody. After washing away any unbound substances, added antibody-conjugate specific for Alpha 1 Antichymotrypsin to each well and incubate. After washing away any unbound substances, the TMB Substrate is added to the wells and color develops in proportion to the amount of Alpha 1 Antichymotrypsin bound in the initial step. The color development is stopped by the addition of

Stop Solution and the intensity of the color is measured at a wavelength of 450 nm. The concentration of Alpha 1 Antichymotrypsin in the samples is then determined by comparing the O.D of samples to the standard curve.

MATERIALS PROVIDED & STORAGE INFORMATION

Store all other components at 2-8°C. Use the kit before expiration date.

Component	Quantity	Storage information
Antibody Coated Microplate	8 x 12 strip	4°C
Standards (lyophilized)	1 vial	4°C
100X Antibody Conjugate	150 μL	4°C (protect from light)
20X Wash Buffer	50 mL	4°C
5X Diluent Buffer	50 mL	4°C
TMB Substrate	12 mL (ready to use)	4°C (protect from light)
Stop Solution	12 mL (ready to use)	4°C

MATERIALS REQUIRED BUT NOT PROVIDED

- Microplate reader capable of reading at 450 nm
- Deionized or distilled water
- Pipettes and pipette tips
- Multichannel micropipette reservoir
- Microtiter plate washer (recommended)

TECHNICAL NOTES AND PRECAUTIONS

- Wear protective gloves, clothing, eye, and face protection especially while handling blood or body fluid samples.
- Store the kit at 2-8°C at all times.
- Return any unused microplate strips to the plate pouch with desiccant.
- Prior to beginning the assay procedure, bring all reagents and required number of strips to room temperature (16-25°C).
- The reagent preparation method might be different from lot to lot, so please check the lot and follow the instructions given in this manual.
- Avoid air bubbles in the wells as this could result in lower binding efficiency and higher CV% of duplicate reading.
- Briefly spin down the all vials before use.
- If crystals are observed in the 20X Wash Buffer, warm to 37°C until the crystals are completely dissolved.
- Minimize lag time between wash steps to ensure the plate does not become completely dry during the assay.
- Ensure complete reconstitution and dilution of reagents prior to use.

- Take care not to contaminate the TMB Substrate. Do not expose the TMB solution to glass, foil or metal. If the solution is blue before use, DO NOT USE IT.
- Do NOT return leftover TMB Substrate to bottle. Do NOT contaminate the unused TMB Substrate. If the solution is blue before use, DO NOT USE IT.
- Change pipette tips between the addition of different reagent or samples.
- Taping the well strips together with lab tape can be done as an extra precaution to avoid plate strips from coming loose during the procedure.
- Run both standards and samples in at least duplicates (triplicate is recommended).

SAMPLE COLLECTION & STORAGE INFORMATION

The following recommendations are only guidelines and may be altered to optimize or complement the user's experimental design.

Serum: Collect blood in a tube with no anticoagulant. Allow the blood to clot at room temperature for 30 minutes. Centrifuge at 1,000 x g for 15 minutes at 4°C. Recommended starting dilution is 1/5,000. To prepare a 1/5,000 dilution of a sample, transfer 2 μL of sample to 198 μL of 1X Diluent Buffer. This gives you a 1/100 dilution. Next, dilute the 1/100 by transferring 6 μL into 294 μL of 1X Diluent Buffer. This gives you a 1/5,000 dilution Mix thoroughly at each stage. Plasma: Collect blood with EDTA or citrate and centrifuge at 1,000 x g for 15 minutes at 4°C. Recommended starting dilution is 1/5,000. To prepare a 1/5,000 dilution of a sample, transfer 2 μL of sample to 198 μL of 1X Diluent Buffer. This gives you a 1/100 dilution. Next, dilute the 1/100 by transferring 6

 μL into 294 μL of 1X Diluent Buffer. This gives you a 1/5,000 dilution Mix thoroughly at each stage.

Note:

- 1. Do not use haemolytic, icteric or lipaemic specimens.
- 2. Avoid disturbing the white buffy layer when collection serum/plasma sample.
- 3. Azide and thimerosal at concentrations higher than 0.1% inhibits the enzyme reaction.
- 4. Store all samples on ice after preparation and use immediately or aliquot and store at-80°C (preferably) or -20°C. Avoid repeated freeze-thaw cycles.

REAGENT PREPARATION

- **1X Wash Buffer**: Dilute 20X Wash Buffer into distilled water to yield 1X Wash Buffer. (E.g., add 50 mL of 10X Wash Buffer into 950 mL of distilled water to a final volume of 1 L)
- 1X Diluent Buffer: Dilute 5X Diluent Buffer into distilled water to yield 1X
 Diluent Buffer. (E.g., add 50 mL of 5X Diluent Buffer into 200 mL of distilled
 water to a final volume of 250 mL)
- 1X Antibody Conjugate: Dilute 100X Antibody Conjugate into 1X Diluent Buffer to yield 1X Antibody Conjugate. (E.g., add 10 μ L of 100X Antibody Conjugate into 990 μ L of 1X Diluent Buffer to a final volume of 1 mL)
- Standards: Prepare according to the Certificate of Analysis. Dilute the Standards in 400, 200, 100, 50, 25, 12.5, 6.25, 0 ng/mL with 1X Diluent Buffer.

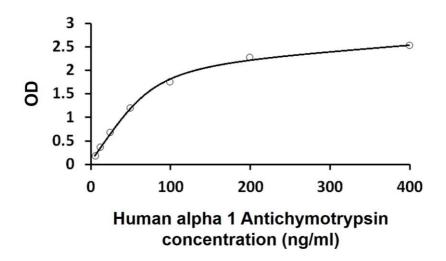
ASSAY PROCEDURE

All materials should be equilibrated to room temperature (RT, 16-25°C) before use. Standards and samples should be assayed in duplicates.

- 1. Add $100 \, \mu L$ of samples and each diluted Standards to the Antibody Coated microplate. Cover the plate with the Plate Sealer. Incubate for $1 \, hour$ at room temperature.
- 2. Aspirate each well and wash, repeating the process 3 times for a total 4 washes. Wash by filling each well with **1X Wash Buffer (300 μL)** using a squirt bottle, manifold dispenser. Complete removal of liquid at each is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating, decanting or blotting against clean paper towels.
- 3. Add **100 μL** of the **1X Antibody Conjugate** per well. Then cover the plate with the Plate Sealer. Incubate for **15 minutes** at **room temperature** in the dark.
- 4. Aspirate and wash plate as in step 2.
- 5. Add $100 \,\mu\text{L}$ of TMB Substrate to each well. Incubate for 5 minutes at room temperature in the dark.
- 6. Immediately Add 100 μ L of Stop solution to each well. The color of the solution should change from blue to yellow.
- 7. Read the absorbance with a microplate reader at **O.D. 450 nm** within **30** minutes.

EXAMPLE OF TYPICAL STANDARD VALUES

The following figures demonstrate typical results with the Human alpha 1 Antichymotrypsin ELISA Kit. One should use the data below for reference only. This data should not be used to interpret actual results.



CALCULATION OF RESULTS

- Subtract zero point (S0) from all standards and unknowns to determine corrected absorbance.
- 2. Using log-log, semi-log or linear graph paper, construct a standard curve by plotting the mean absorbance obtained from each standard against its concentration with absorbance value on the vertical (Y) axis and concentration on the horizontal (X) axis.
- 3. Automated method: The results in the IFU have been calculated

automatically using a 4 PL (4 Parameter Logistics) curve fit. 4 Parameter Logistics is the preferred method. Other data reduction functions may give slightly different results.

- 4. arigo provides GainData®, an in-house development ELISA data calculator, for ELISA data result analysis. Please refer our GainData® website for details. (https://www.arigobio.com/elisa-analysis)
- 5. If the samples have been diluted, the concentration read from the standard curve must be further converted by the appropriate dilution factor according to the sample preparation procedure as described above.

QUALITY ASSURANCE

Sensitivity

2.518 ng/mL

Precision

Intra-CV=3.293%; Inter-CV=3.618%