

# **Lipoxygenase Assay Kit**

ARG83421 Lipoxygenase Assay Kit can be used to measure Lipoxygenase in tissue extracts, cell lysate, cell culture media and other biological fluids.

Catalog number: ARG83421

Package: 96 wells

For research use only. Not for use in diagnostic procedures.

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#### INTRODUCTION

Lipoxygenases (LOXs) are dioxygenases that catalyze the formation of corresponding hydroperoxides from polyunsaturated fatty acids such as linoleic acid and arachidonic acid.

### PRINCIPLE OF THE ASSAY

Lipoxygenase Assay Kit determined Lipoxygenase activity in Tissue extracts, cell lysate, cell culture media and other biological fluids. Lipoxygenase Assay Kit is based on oxidation of substrate to iodine. The Lipoxygenase activity can be measured at a colorimetric readout at 470 nm

#### **MATERIALS PROVIDED & STORAGE INFORMATION**

Component	Quantity	Storage
Microplate	1 X 96-well plate	
Standard (3 μmol/mL)	1 ml	4°C
Assay Buffer	4 X 30 ml	4°C
Reaction Buffer	7 ml	4°C
Substrate I	1 vial (lyophilized)	4°C
Substrate I Diluent	6 ml	4°C
Substrate II	1 vial (lyophilized)	4°C
Reagent Dye	1 ml	4°C

## **MATERIALS REQUIRED BUT NOT PROVIDED**

- Microplate reader capable of measuring absorbance at 470 nm
- Pipettes and pipette tips
- Deionized or distilled water
- If precipitate are observed in the Reagent Dye, warm to 80°C until the precipitate are completely dissolved.

#### **TECHNICAL HINTS AND PRECAUTIONS**

- Wear protective gloves, clothing, eye, and face protection especially while handling blood or body fluid samples.
- Store all component at 4°C.
- Briefly spin down the reagents before use.
- It is highly recommended that the standards and samples be assayed in at least duplicates.
- Change pipette tips between the addition of different reagent or samples.

#### SAMPLE COLLECTION & STORAGE INFORMATION

The sample collection and storage conditions listed below are intended as general guidelines. Sample stability has not been evaluated.

<u>Cell and bacteria</u> - Collect cell or bacteria into centrifuge tube, discard the supernatant after centrifugation, add 1 ml Assay Buffer for 5×106 cell or bacteria, sonicate (with power 20%, sonicate 3s, interval 10s, repeat 30 times); centrifuged at 8000g 4 °C for 10 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

<u>Tissue</u> - Weigh out 0.1 g tissue, homogenize with 1 ml Assay Buffer on ice, centrifuged at 8000g 4 °C for 10 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

Note: For liquid sample, it can be assayed directly.

#### REAGENT PREPARATION

- Substrate I: Reconstitute the Substrate with 6 ml of Substrate I Diluent.
   Allow the Substrate I keep on bench for few minutes. Make sure the Substrate is dissolved completely and mixed thoroughly before use.
- Substrate II: Reconstitute the Substrate with 3 ml of Distilled water. Allow the Substrate II keep on bench for few minutes. Make sure the Substrate is dissolved completely and mixed thoroughly before use.
- **Standard:** Perform 2-fold serial dilutions of the top standards to make the standard curve

#### **ASSAY PROCEDURE**

Standards and samples should be assayed in at least duplicates.

- 1. Add **60 μl** of **Substrate I** into each well.
- 2. Add **30 μl** of **Sample** into Sample well.
- 3. Add **30 μl** of **Standard** into Standard well.
- 4. Add **30 μl** of **Distilled water** into Blank well.
- 5. Mix well. Incubate at 30°C for 5 min.
- 6. Add **70 μl** of **Reaction Buffer** into each well.
- 7. Add **30 μl** of **Substrate II** into each well.
- 8. Add 10 μl of Reagent Dye into each well.
- 9. Mix well. Incubate at RT for 5 min. Read the OD at 470 nm.

## Summary of Lipoxygenase Assay Procedure

Reagent	Sample	Standard	Blank	
Substrate I	60 μl	60 μl	60 µl	
Sample	30 μΙ	-	-	
Standard	-	30 μΙ	-	
Distilled water	-	-	30 μΙ	
Mix well. Incubate at 30°C for 5 min				
Reaction Buffer	70 μΙ	70 μΙ	70 μΙ	
Substrate II	30 μΙ	30 μΙ	30 μΙ	
Reagent Dye	10 μΙ	10 μΙ	10 μΙ	
Mix well. Incubate at RT for 5 min				
Read the OD at 470 nm.				

#### **CALCULATION OF RESULTS**

- 1. Unit Definition: One unit Lipoxygenase activity is defined as the generates one  $\mu$ mol of hydrogen peroxide per minute in the reaction system.
- 2. Calculate the average absorbance values for each set of samples and control.

#### 3. Calculation:

#### A. Definition:

C<sub>Protein</sub>: the protein concentration, mg/ml;

C<sub>Standard</sub>: the concentration of Standard, 3 µmol/ml;

 $V_{Sample}$ : the volume of reaction sample, 30  $\mu$ l = 0.03 ml;

 $V_{Standard}$ : the volume of reaction Standard, 30  $\mu$ l = 0.03 ml;

 $V_{assay}$ : the volume of Assay Buffer, 1000  $\mu$ l = 1 ml;

W: the weight of sample, g;

N: the quantity of cell or bacteria,  $N \times 10^4$ ;

T: the reaction time, 5 minutes.

#### B. Formula:

a). According to the protein concentration of sample

V<sub>Sample</sub>) X T]

= 0.6 X (OD<sub>Sample</sub> - OD<sub>Blank</sub>) / [(OD<sub>Standard</sub> - OD<sub>Blank</sub>) X C<sub>Protein</sub>]

b). According to the weight of sample

$$\label{eq:lipoxygenase} \begin{tabular}{ll} Lipoxygenase activity (U/mg) = & $(OD_{Sample} - OD_{Blank})$ X (C_{Standard} X V_{Standard}) / [(OD_{Standard} - OD_{Blank}) X (W X V_{Sample} / V_{Assay}) X T] \end{tabular}$$

= 0.6 X (ODsample - ODBlank) / [(ODstandard - ODBlank) X W]

c). According to the cell or bacteria

Lipoxygenase activity 
$$(U/10^4) = (OD_{Sample} - OD_{Blank}) \times (CS_{Standard} \times VS_{Sample}) / [(OD_{Standard} - OD_{Blank}) \times (N \times VS_{Sample}) / VASSAY) \times T]$$

$$= 0.6 \times (OD_{Sample} - OD_{Blank}) / [(OD_{Standard} - OD_{Blank}) \times N]$$

#### **EXAMPLE OF TYPICAL RESULT**

The following data is for demonstration only and cannot be used in place of data generations at the time of assay. Please note this data is for demonstration only and serially diluted standards are necessary for this kit.

