



# CA / Carbonic Anhydrase Activity Assay Kit

ARG83557 CA / Carbonic Anhydrase Activity Assay Kit can be used to measure CA / Carbonic Anhydrase in Serum, Plasma, Tissue extracts, Cell lysate, Cell culture media, other biological fluids

Catalog number: ARG83557

Package: 96 wells

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For research use only. Not for use in diagnostic procedures.

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## CA / Carbonic Anhydrase Activity Assay Kit ARG83557

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### PRINCIPLE OF THE ASSAY

ARG83557 CA / Carbonic Anhydrase Activity Assay Kit provides a simple and sensitive method for monitoring Carbonic Anhydrase activity in various samples. The enzyme catalyzed reaction products p-nitrophenol can be measured at a colorimetric readout at 405 nm.

### MATERIALS PROVIDED & STORAGE INFORMATION

Store Standard and Positive Control at -20 °C, all other component at 2-8°C.

Use the kit before expiration date.

| Component            | Quantity             | Storage              |
|----------------------|----------------------|----------------------|
| Microplate           | 1 X 96-well plate    |                      |
| Standard (2 µmol/mL) | 1 ml                 | -20 °C, keep in dark |
| Substrate            | 2 vial (lyophilized) | 4 °C, keep in dark   |
| Substrate Diluent    | 0.3 ml               | 4 °C                 |
| Reaction Buffer      | 25 ml                | 4 °C                 |
| Positive Control     | 1 vial (lyophilized) | -20 °C               |
| Assay Buffer         | 4 X 30 ml            | 4 °C                 |
| Plate sealer         | 3 adhesive strips    |                      |

### MATERIALS REQUIRED BUT NOT PROVIDED

- Microplate reader capable of measuring absorbance at 405 nm
- Pipettes and pipette tips
- Deionized or distilled water

### TECHNICAL HINTS AND PRECAUTIONS

- Wear protective gloves, clothing, eye, and face protection especially while handling blood or body fluid samples.
- Store Standard and Positive Control at -20 °C, all other component at 2-8°C. Use the kit before expiration date.
- Briefly spin down the reagents before use.
- It is highly recommended that the standards and samples be assayed in at least duplicates.
- Change pipette tips between the addition of different reagent or samples.

### SAMPLE COLLECTION & STORAGE INFORMATION

The sample collection and storage conditions listed below are intended as general guidelines. Sample stability has not been evaluated.

**Cell and Bacteria** - Collect cell or bacteria into centrifuge tube, discard the supernatant after centrifugation. Mix and sonicate (with power 20%, sonicate 3s, interval 10s, repeat 30 times) with 1 ml Assay buffer per  $5 \times 10^6$  cell or bacteria. Centrifuged at 10,000g 4 °C for 20 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

**Tissue** - Weigh out 0.1 g tissue, homogenize with 1 ml Assay buffer on ice, centrifuged at 10,000g 4 °C for 20 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

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**Liquid samples** - Detect directly.

**Serum** - Use a serum separator tube (SST) and allow samples to clot for 30 minutes before centrifugation for 15 minutes at 1000 x g. Collect serum and assay immediately or aliquot & store samples at -20°C up to 1 month or -80°C up to 6 months. Avoid repeated freeze-thaw cycles.

**Plasma** - Collect plasma using heparin as an anticoagulant. Centrifuge for 15 minutes at 1000 x g. within 30 minutes of collection. Collect the supernatants and assay immediately or aliquot and store samples at -20°C up to 1 month or -80°C up to 6 months. Avoid repeated freeze-thaw cycles.

### REAGENT PREPARATION

- **Substrate:** Briefly centrifuge prior to opening. Dissolve each substrate vial in **0.15 ml** Substrate Diluent before use. Make sure the Substrate is dissolved completely and mixed thoroughly before use. Keep in dark and store at -20 °C for 7 days or 4°C for 12 hours
- **Substrate Diluent:** Thaw Substrate Diluent at room temperature before use.
- **Substrate Working Solution:** Prepare ready-to-use Substrate Working Solution by mixing Reaction Buffer and Substrate at a ratio of 187:3 (v/v). Vortex thoroughly until no precipitate is visible and use immediately.
- **Positive Control:** Reconstitute the **Positive Control** with **1 ml** of Assay Buffer. Allow the **Positive Control** keep on bench for few minutes. Make sure the **Positive Control** is dissolved completely and mixed thoroughly before use. The **Positive Control** is stable for up to 4 weeks at -80°C. Mix well before use.
- **Standard:** Completely dissolved and briefly centrifuge prior to opening.

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Perform 2-fold serial dilutions of the top standard solution using Reaction Buffer to make the standard curve. The concentration of standard curve could be 2  $\mu\text{mol/mL}$ , 1  $\mu\text{mol/mL}$ , 0.5  $\mu\text{mol/mL}$ , 0.25  $\mu\text{mol/mL}$ , 0.125  $\mu\text{mol/mL}$ , 0.063  $\mu\text{mol/mL}$ , 0.031  $\mu\text{mol/mL}$ , 0.016  $\mu\text{mol/mL}$ . Keep in dark and store at  $-20\text{ }^{\circ}\text{C}$  for 1 month or  $4\text{ }^{\circ}\text{C}$  for 3 days.

Note: Divide into small aliquots to avoid repeated freeze-thaw cycles.

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### ASSAY PROCEDURE

Standards and samples should be assayed in at least duplicates.

1. Add **190  $\mu$ l** Substrate Working Solution into **Sample, Control** and **Positive Control wells**.
2. Add **180  $\mu$ l** Reaction Buffer into **Standard** and **Blank wells**.
3. Add **10  $\mu$ l** Distilled water into each **Control wells**.
4. Add **20  $\mu$ l** Distilled water into each **Blank wells**.
5. Add **20  $\mu$ l** Standard into each **Standard wells**.
6. Add **10  $\mu$ l** Positive Control into each **Positive Control wells**.
7. Add **10  $\mu$ l** Sample into **Sample wells**.
8. Mix well. Put in the oven, **incubate** at 37 °C for **5 min**. Read the OD at **405 nm**.

#### Summary of CA / Carbonic Anhydrase Activity Assay Kit Procedure

| Reagent   | Sample      | Control     | Positive Control | Standard    | Blank       |
|---|-------------|-------------|------------------|-------------|-------------|
| Substrate Working Solution  | 190 $\mu$ l | 190 $\mu$ l | 190 $\mu$ l      |             |             |
| Reaction Buffer   |             |             |                  | 180 $\mu$ l | 180 $\mu$ l |
| Distilled water   | -           | 10 $\mu$ l  | -                | -           | 20 $\mu$ l  |
| Standard  | -           | -           | -                | 20 $\mu$ l  | -           |
| Positive Control  | -           | -           | 10 $\mu$ l       | -           | -           |
| Sample  | 10 $\mu$ l  | -           | -                | -           | -           |
| Mix well. Put in the oven, <b>incubate</b> at 37 °C for <b>5 min</b> . Read the OD at <b>405 nm</b> . |             |             |                  |             |             |

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Note:

1. Reagents must be added sequentially and should not be premixed prior to addition.
2. For unknown samples, we recommend doing a pilot experiment & testing several doses to ensure the readings are within the standard curve range. If the enzyme activity is lower, please add more samples into the reaction system; or increase the reaction time; if the enzyme activity is higher, please dilute the sample, or decrease the reaction time.

### CALCULATION OF RESULTS

1. Calculate the average absorbance values for each set of samples, standard and blank.

a.) Definition: One unit of Carbonic Anhydrase activity is defined as the enzyme generates 1 nmol of p-nitrophenol per minute.

$C_{\text{Standard}}$ : the concentration of standard, 2  $\mu\text{mol/mL}$  =2000 nmol/ml;

$C_{\text{Protein}}$ : the protein concentration, mg/ml;

W: the weight of sample, g;

N: the quantity of cell or bacteria,  $N \times 10^4$ ;

$V_{\text{Standard}}$ : the volume of the standard, 0.02 ml;

$V_{\text{Sample}}$ : the volume of sample, 0.01 ml;

$V_{\text{Assay}}$ : the volume of Assay buffer, 1 ml;

T: the reaction time, 5 minutes.

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According to the slope of the standard curve

Activity =

$$\{[(\text{OD}_{\text{Sample}} - \text{OD}_{\text{Control}}) / - \text{Intercept}] / \text{Slope} \times T\} \times n \text{ (U/mL)}$$

Calculate the initial activity according to sample preparation procedure.

b.) Calculation:

Formula:

a). According to the protein concentration

$$\text{CA (U/mg)} =$$

$$(\text{C}_{\text{Standard}} \times \text{V}_{\text{Standard}}) \times (\text{OD}_{\text{Sample}} - \text{OD}_{\text{Control}}) / [(\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) \times (\text{C}_{\text{Protein}} \times \text{V}_{\text{Sample}}) \times T]$$

$$= 800 \times (\text{OD}_{\text{Sample}} - \text{OD}_{\text{Control}}) / [(\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) \times \text{C}_{\text{Protein}}]$$

b). According to the weight

$$\text{CA (U/mg)} =$$

$$(\text{C}_{\text{Standard}} \times \text{V}_{\text{Standard}}) \times (\text{OD}_{\text{Sample}} - \text{OD}_{\text{Control}}) / [(\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) \times (\text{V}_{\text{Sample}} \times \text{W} / \text{V}_{\text{Assay}}) \times T]$$

$$= 800 \times (\text{OD}_{\text{Sample}} - \text{OD}_{\text{Control}}) / [(\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) \times \text{W}]$$

c). According to the quantity of cell or bacteria

$$\text{CA (U/10}^4\text{)} =$$

$$(\text{C}_{\text{Standard}} \times \text{V}_{\text{Standard}}) \times (\text{OD}_{\text{Sample}} - \text{OD}_{\text{Control}}) / [(\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) (\text{V}_{\text{Sample}} \times \text{N} / \text{V}_{\text{Assay}}) \times T]$$

$$= 800 \times (\text{OD}_{\text{Sample}} - \text{OD}_{\text{Control}}) / [(\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) \times \text{N}]$$

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4. According to the volume

CA (U/mg) =

$$\frac{(C_{\text{Standard}} \times V_{\text{Standard}}) \times (OD_{\text{Sample}} - OD_{\text{Control}})}{[(OD_{\text{Standard}} - OD_{\text{Blank}}) \times V_{\text{Sample}} \times X \text{ T}]}$$

$$= 800 \times (OD_{\text{Sample}} - OD_{\text{Control}}) / (OD_{\text{Standard}} - OD_{\text{Blank}})$$

Assay range: 0.016- 2  $\mu\text{mol/mL}$

Sensitivity: 0.016  $\mu\text{mol/mL}$