

Tryptophan Assay Kit

ARG83608 Tryptophan Assay Kit is a detection kit for the quantification of Tryptophan.

Catalog number: ARG83608

Package: 100 assays

For research use only. Not for use in diagnostic procedures.

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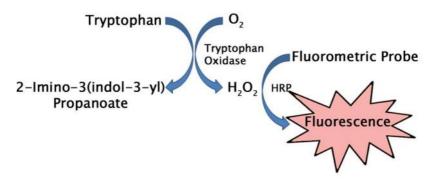
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PRINCIPLE OF THE ASSAY

ARG83608 Tryptophan Assay Kit is a sensitive quantitative fluorometric assay for tryptophan. Tryptophan Oxidase converts tryptophan to 2-imino-3(indol-3-yl) propanoate and hydrogen peroxide (H2O2). The H2O2 is then detected with a highly specific fluorometric probe. Horseradish peroxidase catalyzes the reaction between the probe and hydrogen peroxide, which bind in a 1:1 ratio. Samples and standards are incubated for 60 minutes and then read with a standard 96-well fluorometric plate reader. Samples are compared to a known concentration of tryptophan standard within the 96-well microtiter plate format.



MATERIALS PROVIDED & STORAGE INFORMATION

Upon receipt, Tryptophan Oxidase at -80°C, other component store at -

20°C. Use the kit before expiration date.

Component	Quantity	Storage information
L-Tryptophan Standard (<u>10 mM</u>)	100 μΙ	-20°C
Tryptophan Oxidase	500 μΙ	-20°C
Probe	50 μΙ	-20°C
HRP	100 μΙ	RT
10X Assay Buffer	30 ml	RT

MATERIALS REQUIRED BUT NOT PROVIDED

- Black microplate reader
- Pipettes and pipette tips
- Deionized or distilled water
- Centrifuge spin filter

TECHNICAL HINTS AND PRECAUTIONS

- Wear protective gloves, clothing, eye, and face protection especially while handling blood or body fluid sample.
- Upon receipt, Tryptophan Oxidase at -80°C, other component store at -20°C. Use the kit before expiration date.
- All reagents should be mixed by gentle inversion or swirling prior to use.
 Do not induce foaming.
- Before using the kit, spin tubes and bring down all components to the bottom of tubes.
- Change pipette tips between the addition of different reagent or sample.

SAMPLE COLLECTION & STORAGE INFORMATION

The sample collection and storage conditions listed below are intended as general guidelines. Sample stability has not been evaluated.

Cell Culture Supernatants- Remove particulates by centrifugation for 10 min at 1500 x g at 4°C and aliquot & store samples at-20°C up to 1 month or-80°C up to 6 months. Avoid repeated freeze-thaw cycles.

Serum- Use a serum separator tube (SST) and allow samples to clot for 30 minutes before centrifugation for 15 minutes at 1000 x g. Collect serum and assay immediately or aliquot & store samples at-20°C up to 1 month or-80°C up to 6 months. Avoid repeated freeze-thaw cycles.

Plasma- Collect plasma using EDTA or heparin as an anticoagulant. Centrifuge for 15 minutes at 1000 x g. within 30 minutes of collection. Collect the supernatants and assay immediately or aliquot and store samples at-20°C up to 1 month or-80°C up to 6 months. Avoid repeated freeze-thaw cycles.

Urine- Collect the first urine of the day, micturate directly into a sterile container. Remove impurities by centrifugation. Collect the supernatants and assay immediately or aliquot and store samples at \leq -20°C. Avoid repeated freeze-thaw cycles.

Cell or Tissue Lysate- Sonicate or homogenize sample in cold PBS and centrifuge at 10,000 x g for 10 minutes at 4°C. Collect samples and assay immediately or aliquot and store samples at-80°C. Avoid repeated freeze-thaw cycles.

REAGENT PREPARATION

- 1x Assay Buffer Dilute the 10x Assay Buffer into Deionized Water to yield 1x Assay Buffer. The 1x Assay Buffer is stable for up to 6 months at 2-8°C.
- Reaction / Negative Control Mix: Prepare as below:

Component	Reaction Mix	Negative Control Mix
Fluorometric Probe	25 μΙ	25 μΙ
HRP	5 μΙ	5 μΙ
Tryptophan Oxidase	250 μΙ	
1X Assay Buffer	2220 μΙ	2470 μΙ
Total	2500 μΙ	2500 μΙ

Standards: Prepare fresh Lysine Standards before use by diluting in 1X
 Assay Buffer according to the Table below.

Standard	Tryptophan	1X Assay Buffer	Standard
		·	
tube	(μM)	(μL)	(μL)
64	S1 100 495	405	5
51		495	(1 <u>0 mM</u> Tryptophan)
S2	50	250	250 of S1
S3	25	250	250 of S2
S4	12.5	250	250 of S3
S5	6.25	250	250 of S4
S6	3.13	250	250 of S5
S7	1.56	250	250 of S6
S0	0	200	0

ASSAY PROCEDURE

All materials should be equilibrated to room temperature (RT, 20-25°C) before use. Standards and sample should be assayed in duplicates.

- 1. Add $50 \,\mu l$ of diluted sample or each diluted Standard into respective wells of the 96-well plate.
- 2. Add **50 μl** of **Reaction / Negative Control Mix** to each well.
- 3. Cover the plate and incubate for **60 minutes** at **RT**.
- 4. Read the absorbance with a plate reader at O.D. 530-570 nm.

CALCULATION OF RESULTS

Plot the RFU measured at 60 minutes for each standard against the standard concentrations. Determine the slope using linear regression fitting.

The Tryptophan concentration of a sample is calculated as follow:

Net RFU =
$$(RFU_{+TO})$$
- (RFU_{-TO})

EXAMPLE OF TYPICAL STANDARD CURVE

The following figures demonstrate typical results with the Tryptophan. One should use the data below for demonstration only and cannot be used in place of data generations at the time of assay.

